171. The Structure of Covalent Hydrates¹) of Alloxazines²)

A Reinvestigation

by Jacek Koziol³) and Bozena Tyrakowska

Institute of Commodity Sciences, Academy of Economics, ul. Marchlewskiego 146/150, 60-967 Poznan, Poland

and Franz Müller

Department of Biochemistry, Agricultural University, De Dreijen 11, 6703 BC Wageningen, The Netherlands

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Summary

It is shown by ¹³C-NMR. and synthesis of 6, 7-dimethyl-3-ethoxycarbonylaminoquinoxaline-2-carboxamide (5) and methyl 6, 7-dimethyl-3-(N'-methylureido)quinoxaline-2-carboxylate (6) that hydrolysis of lumichrome (1, R = H) must yield 6, 7-dimethyl-3-ureidoquinoxaline-2-carboxylic acid (2, R = H) and not 6, 7-dimethyl-2-oxo-1,2,9,9a-tetrahydrooxazolo[4,5-b]quinoxaline-9a-carboxamide (2a, R = H), as previously proposed.

Introduction. - As mentioned in the literature [1] [2], alloxazines unsubstituted at N(1) undergo a reversible reaction in aqueous alkaline solution yielding 'covalent hydrates'. For the product obtained from lumichrome⁴) the structure **2a** ($\mathbf{R} = \mathbf{H}$) (cf. Scheme) has been proposed [3]. More recent data [4], however, seem inconsistent with the proposed structure. Therefore, the structure of the 'covalent hydrates' was reinvestigated using ¹³C-NMR. and chemical synthesis. We show that the correct structure of the 'covalent hydrate' of alloxazine is 6,7-dimethyl-3-(N'-methylureido)quinoxaline-2-carboxylic acid (**2**, $\mathbf{R} = \mathbf{CH}_3$).

Results and discussion. – For the unambiguous synthesis of the 'covalent hydrate' two routes were chosen, with methyl 3-amino-6, 7-dimethyl-quinoxaline-2-carboxylate (3) (cf. Scheme) as the starting material. The ester 3 was used because the corresponding free acid undergoes easy decarboxylation under the conditions of the synthesis. The free acid corresponding to 3 is, in fact, a product of exhaustive hydrolysis of lumichromes (1, R = H or CH_3) [8]. The aim of the synthesis of 5 and 6 was to compare the physical and chemical properties of these compounds

¹) For the term 'covalent hydrates', see [1].

²) Alloxazine = 1H, 3H-benzo[g]pteridine-2, 4-dione.

³⁾ To whom correspondence should be addressed.

⁴) Lumichrome = 7, 8-dimethylalloxazine.



with those of the esters of the 'covalent hydrates' of 1 for which structure 2b was proposed. Compound 5 but not 6 should be identical with 2b (R=H, $R'=C_2H_5$).

The syntheses of 5 and 6 were similar to the procedures described by *Clerin et al.* [12] for the preparation of isoalloxazine (= flavin).

a) A solution of 3 in CH₃NCO yields 6 in a slow reaction and also some 1 ($R = CH_3$). Compound 6 possesses the same physical and chemical properties (*cf. Table 1*) as the methyl ester of 2 ($R = CH_3$) and can be converted to 1 ($R = CH_3$) in aqueous solutions of pH > 6 or in methanolic KOH-solution.

b) The second synthesis consisted of the transformation of 3 into 4 by ethyl chloroformate. The product 4 was in turn converted to the amide 5 which is different from the ethyl ester of the 'covalent hydrate' [3] with R = H (see *Table 1*). The conditions for the conversion of 5 into 1 (R = H) are less mild than those for the conversion of $6 \rightarrow 1$ ($R = CH_3$).

From these results it follows that 2 is the true structure of the 'covalent hydrate'.

This structure was confirmed by ¹³C-NMR. measurement of selectively ¹³Cenriched compounds. The most significant difference between 2 and 2a is the state of hybridization of the C(4a)-atom (after numbering of flavins). Therefore, the chemical shift of C(4a) provides independent and unequivocal evidence for the real structure. The results are collected in *Table 2*. The chemical shifts are in agreement with structure 2 for the 'covalent hydrate' but not with the earlier proposed structure 2a. In the latter case the resonance signal of C(4a) would be expected at much higher field, similar to that of a C(4a)-substituted flavin [13]. The normalized intensities of the signals of proton noise decoupled spectra (*Table 2*) also agree with structure 2 (R = CH₃), but not with structure 2a (R = CH₃).

	Table 1. Comparison of some	physical data of 5, 6 and methyl	and ethyl ester of 2	
Technique	Compound			
	Q	2 (methyl ester) $(R = CH_3)^a$)	S	2 (ethyl ester) (R = H) ^b)
UV. (methanol)	376 nm (3.70) 334 nm (3.78)	376 nm (3.70) 331 nm (3.78)	365 nm (3.83) 370 nm (3.80)	378 nm (3.76) 318 nm (3.83)
<i>λ</i> .max (108ε) IR. ŷ(NH) [cm ⁻¹]	3320, 3270	3320, 3260	3460, 3260	3310, 3180
IR. v(CO) [cm ⁻¹]	1687	1690	1740, 1675	1725, 1695
¹ H-NMR. ^c)	2.44 (CH ₃ C(7));	2.44 (CH ₃ C(7));	1.5-1.3 (CH ₃) ^f);	1.55-1.40 (CH ₃) ^f);
×	2.50 (CH ₃ C(6));	2.52 (CH ₃ C(6));	2.45 (CH ₃ C(7));	2.40 (CH ₃ C(7));
CDC1,	$3.07-2.98^{d}$) (CH ₃ N(3));	$3.10-3.00 (CH_3N(3));$	2.55 (CH ₃ C(6));	2.45 (CH ₃ C(6));
1	4.05 (CH ₃ O);	4.05 (CH ₃ O);	4.6-4.2 (CH ₂) ^g);	4.65-4.45 (CH ₂) ^g);
	7.52 (H–C(5));	7.57 (H–C(5));	7.5 (NH ₂);	7.55 (H–C(5));
	7.81 (H–C(8));	7.83 (H–C(8));	7.85 (H–C(5));	7.85 (H–C(8));
	9.3 $(H-N(3))^{e}$;	9.21 $(H-N(3))^{e}$;	8.00 (H-C(8));	10.28 (H-N(3)
	10.12 (H-N'(3)) ^e)	$10.10 (H-N'(3))^{e}$	8.40 (H-N(3)	and $H-N'(3))^{e}$
			and $H-N'(3))^{e}$	
^{a)} Published data [3] for the propulation of t	osed structure 2b ($\mathbf{R} = \mathbf{R}' = \mathbf{CH}_3$) c system (cf. Scheme). ^d) d, $J = 5$. ^b) According to [3] this comp 5 Hz, proton exchangeable with	bound possesses structure 2b (R) deuteron (3.03 ppm) . ^c) <i>m</i> , p	= H, R'= C ₂ H ₅). ^c) Ring proton exchangeable with
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Compound	Solvent	Chemical Shifts (Intensities)			
		C(2)	C(4)	C(4a)	C(10a)
$\overline{1 \ (\mathbf{R} = \mathbf{C}\mathbf{H}_3)^{\mathbf{a}})}$	DMSO	150.35 (101)	160.61 ^b) (20)	129.53 (34)	145.39 ^b) (37)
			160.32 ^b) (27)		145.10 ^b) (30)
2 $(R = CH_3)^a)^d$	DMSO	154.22 (140)	167.34°) (25)	131.28 (45)	146.73°) (30)
			167.14°) (30)		146.56°) (35)
6 ^d) ^e)	CDCl ₃	154.71 (120)	165.48 (45)	135.63 (50)	146.53 (100)
NHCH3 djejfj					
с по солнена с	CDCl ₃	-	166.39 (100)	133.03 (60)	152.37 (115)
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Table 2. The relevant ¹³C-chemical shifts (in ppm) and the normalized intensities of the corresponding resonance lines of alloxazines and their 'covalent hydrates'

^a) Compounds selectively ¹³C-enriched, s. experimental part. ^b) d, ² $J_{C(4), C(10a)} = 7.2$ Hz. ^c) d, ² $J_{C(4), C(10a)} = 5.0$ Hz. ^d) Ring numbering is according to that of alloxazine. ^e) Natural abundance spectra. ^f) Prepared as described in [14].

Structure 2 is also in agreement with the published physical and chemical data for the 'covalent hydrate' [3] which, however, have led to the proposal of structure 2a by an incorrect interpretation.

Other observations can now be explained. Thus, N(1)-substituted and N(1), N(3)-unsubstituted alloxazines are very stable towards hydrolysis at high pH values (ca. 14), in contrast to N(1), N(3)-dialkylated compounds which hydrolyze easily to N-alkyl-3-alkylamino-6, 7-dimethyl-quinoxaline-2-carboxamide [14]. This means that whenever the N(3)-position is ionized the carbonyl functions at positions 2 and 4 are protected from attack by hydroxyl ions. Only at pH < pK of N(3) (ca. 12.5 for the dianion and lower for the monoanion) can hydrolysis occur.

On the other hand, N(3)-alkylated alloxazines form the N(1) anion with a pK of 8.50 [5]. The negative charge of this species, mainly localized on the N(1)-C(2)=O region of the molecule, protects C(2)=O but allows attack of C(4)=O by hydroxyl ions and at the same time protects the product from further hydrolysis.

The reversible formation of 1 from 2 is surprising, although precedence for intramolecular nucleophilic attack of an ureido group upon a carboxyl group exists: *Hegarty & Bruice* [15] found that 2-ureido-benzoic acid undergoes a cyclization which is irreversible in weak acid or alkaline aqueous solution. In addition *Smith & Bruice* [16] have reported that alkaline hydrolysis of isoalloxazines (flavins) yields the corresponding, but intermediate product of 2 which reacts reversibly under certain conditions. Therefore it must be concluded that hydrolysis of alloxazines and isoalloxazines follow the same route with respect to the initially formed product.

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Experimental Part

General. UV./VIS. spectra were recorded on a Cary 118c spectrophotometer. IR.-spectra (KBr pellets) were measured on a Specord IR.-75 spectrophotometer. The ¹H-NMR. spectra were taken on a 60 MHz Varian 360 A spectrometer. The ¹³C-NMR. spectra were recorded either on a JEOL FX-60 or on a Varian XL-100 spectrometer operating at 25.2 MHz. All ¹³C-NMR. spectra were acquired in the Fourier transform mode using 12 mm taperlock tubes and under proton noise decoupling conditions. TMS was used as an internal standard. Mass spectra were obtained on a JEOL D-100 spectrometer (m/z values were given).

Melting points (m.p.) are uncorrected. The purity of the compounds was judged by thin layer chromatography (TLC.) (silica gel 1B2, *Bakerflex*) with CHCl₃/2-butanone 14:1 (ν/ν).

Lumichrome (1, R = H) and N(3)-methyllumichrome (1, $R = CH_3$) were prepared according to published procedures [5] [6]. The synthesis of the 'covalent hydrate' of 1 (R = H, CH_3) has been described [3]. Compound 3 was prepared by methylation of 3-amino-6,7-dimethylquinoxaline-2-carboxylic acid [7] obtained according to the procedure of M_cNutt [8].

¹³C-labelled N(3)-methyllumichrome (${}^{13}C_4$ -1, R=CH₃). An equimolar mixture (70 mg) of [4,10a- ${}^{13}C_2$]-, [4a- ${}^{13}C$]- and [2- ${}^{13}C$]-N(3)-methyllumiflavin [9] was dissolved in 1 ml DMF in a test tube, 3 g of urea was added and the mixture heated at 120° for 24 h. After this time the reaction was almost completed (TLC.). The gel-like content of the test tube was then cooled to RT. and added to 10 ml 2 M aqueous acetic acid. The suspension was extracted with CHCl₃, the organic phase thoroughly washed with water and dried (Na₂SO₄). The CHCl₃ phase was evaporated, the residue dissolved in a mixture of CHCl₃/2-butanone 14:1 (ν/ν) and chromatographed through a short column [10]. The fraction containing ${}^{13}C_4$ -1 was evaporated to dryness and the residue crystallized from methanol yielding 41 mg (62%) of pure compound, m.p. 225° ([5]: 225°).

The 'covalent hydrate' of ¹³C₄-1 was prepared as described in [3].

6,7-Dimethyl-3-ethoxycarbonylaminoquinoxaline-2-carboxamide (5). Dry NH₃ was slowly purged into a solution of 100 mg of 4, obtained from 3 [11], in 40 ml of methanol at 24° until 4 no longer was detectable by TLC. The solution was then rapidly evaporated i.v. to dryness at <40°. The crude product thus obtained was twice crystallized from methanol giving 62 mg (65%) of pure 5, yellow needles, m.p. 235°. – MS.: 288 (M^+). The compound possessed a green fluorescence. For other physical data, see *Table 1*.

Methyl 6, 7-dimethyl-3-(N'-methylureido)quinoxaline-2-carboxylate (6). A solution of 40 mg of 3 in 20 ml of freshly distilled methyl isocyanate was kept in the dark at RT. for about 72 h. Some 1 (R=CH₃) was formed besides 6 (TLC.). The reaction mixture was evaporated to dryness. The residue was dissolved in CHCl₃ from which 6 was obtained pure (17 mg, 34%) as pale yellow crystals by the addition of isopropyl ether; m.p. 186°. - MS. (m/z): 288 (M⁺). For other physical data, see Table 1.

Ethyl ester of 'covalent hydrate' was prepared from the 'covalent hydrate' of 1 (R = H) following the procedure for the methyl ester [3]. The crude product was dissolved in a mixture of CHCl₃/2-butanone 14:1 (ν/ν) and chromatographed [10]. The main fraction was collected, evaporated to dryness and the residue crystallized from ethanol as pale yellow crystals (66%); m.p. 224°. - MS.: 288 (M^+). For other physical data see *Table 1*.

Synthesis of lumichrome (1, R=H) and N(3)-methyllumichrome $(1, R=CH_3)$. - a) From 5. To a solution of 100 mg of 5 in 18 ml of methanol was added 2 ml of 1_M methanolic KOH-solution. The reaction is completed in a few hours at RT. (TLC.). The reaction mixture was neutralized with 1_M HCl, the precipitate filtered off and crystallized from methanol. The yield of 1 (R=H) was 70 mg (83%).

b) From 6. To a solution of 100 mg of 6 in 18 ml of methanol was added 2 ml 1M methanolic KOH-solution. The reaction is complete within a few minutes. The precipitate, obtained after neutralization of the reaction mixture with 1M HCl, was collected and crystallized from methanol. The yield of $1 (R = CH_3)$ was 75 mg (84%).

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